

V.V. Shelkovnikov\*, A.M. Altyev, M.S. Fryanova

<sup>1</sup>National Research Tomsk State University, Tomsk, Russian Federation

(\*Corresponding author's e-mail: [shvv@chem.tsu.ru](mailto:shvv@chem.tsu.ru))

## Study of the Methionine Electrooxidation at an Electrode Modified with Vitamin B<sub>12</sub> and Multi-Walled Carbon Nanotubes

The electrochemical behavior of methionine at an electrode modified with vitamin B<sub>12</sub> and multi-walled carbon nanotubes has been studied in this work. A possible mechanism of redox processes occurring on the modified electrode is proposed. When the electrode is formed, vitamin B<sub>12</sub> is fixed on the surface of a carbon-containing electrode modified with multi-walled carbon nanotubes due to adsorption. In the process of cathodic polarization, cobalt(III) in cobalamin is reduced to cobalt(II) to which a partially negatively-charged sulfur atom in the methionine molecule is attached due to electrostatic interactions. During anodic polarization, cobalt(II) in the complex is oxidized up to Co<sup>3+</sup>, the methyl group from methionine is transferred to cobalamin, and homocysteine passes into the solution, which is proved by Raman spectra. Based on the study of the dependence of current and potential on the sweep speed, it was found that the oxidation process is not reversible, the limiting stage is adsorption, and one proton and one electron participate in the electrochemical stage. The following optimal conditions for recording an analytical signal were selected: the background electrolyte is a tartrate buffer solution with pH = 4.01; the electrolysis potential is 1.6 V; the accumulation time is up to 180 s. The metrological characteristics of the procedure for determining methionine were estimated. The accuracy index did not exceed 29 %, the repeatability and intermediate precision indices did not exceed 14 % and 16 %, respectively. The range of detectable contents was (1–50)×10<sup>-7</sup>; the detection limit was 5.0×10<sup>-8</sup> M. It was shown that a 10-fold excess of ascorbic acid, tryptophan, glycine, cysteine and tyrosine does not have a noticeable distorting effect on the procedure for determining methionine.

**Keywords:** methionine, modified electrode, vitamin B<sub>12</sub>, electrooxidation, cobalamin, electron, proton, stripping voltammetry, multi-walled carbon nanotubes.

### Introduction

Methionine is an essential amino acid that plays an important role as a precursor for all other sulfur-bearing amino acids and their derivatives in living organisms. It involves in the regulation of metabolic processes of the immune system and has antioxidant activity. The methionine concentration in the range of 5–15 μmol/l is considered normal for men and women. The increased content of methionine is associated with damage caused to nucleic acids, with various types of cancer and cardiovascular diseases. The reduced methionine concentration leads to neurodegenerative diseases [1–6]. In this regard, the determination of the methionine content in foodstuff and biological fluids is a relevant task. Electrochemical methods of analysis are promising for this purpose.

The mechanism of methionine electrooxidation remains ambiguous. In 1960, dehydro-methionine was proposed in [7] as an intermediate of the product of anodic oxidation of methionine at a platinum wire electrode with pH = 7.0. Dehydro-methionine was also identified as a by-product of methionine oxidation using various analytical methods [8–10]. A few years later, voltammetric determination of methionine at platinum electrodes was reported in [11]. The authors observed a voltammetric wave between +0.7 and +1.0 V (calomel electrode), which manifested itself only in acidic media and only in the presence of hydrogen and chloride ions [11]. Later, owing to the development of new electrode materials, more detailed studies were carried out. In [12] the authors demonstrated methionine oxidation at a graphite electrode, which is an irreversible process controlled by diffusion. In [13], the authors reported that the oxidation potential of methionine at a glassy-carbon electrode remains constant in the pH range from 3 to 7. The authors also noted that the voltammetric wave of methionine oxidation strongly resembles cystine obtained in similar experimental conditions [12] at the glassy-carbon electrode. In 1994, the authors [14] showed that the neighboring carboxylate and amino groups of thioester, which modifies the electrode, play an important role in the methionine elec-

trooxidation at the carbon electrode, even if these groups are not rigidly held in a fixed position relatively the function of thioester in the molecular skeleton. Later, in 2011, glassy-carbon and diamond electrodes doped with boron were used to study the electrochemical oxidation of methionine in a wide pH range using cyclic and differential pulse voltammetry [15]. The results conform fully to the conclusions of the work [13]. In [15] the oxidation reaction proceeds in two stages at the glassy-carbon electrode and in one stage at boron-doped diamond electrodes, where, probably, the high oxidation current of the second peak is superimposed on the first one. Oxidation reactions have been found to be irreversible, diffusion-controlled, and one-electron processes, independent of pH at  $\text{pH} > 3$ .

In [16], we proposed a technique for determining methionine at an electrode modified with cyanocobalamin (vitamin B<sub>12</sub>). When choosing the technique for electrode modification, we proceeded from the following assumptions: methionine entering the human body, when interacting with enzymes, and is reduced to homocysteine [17]. Such reaction can serve as a prototype of the corresponding electrode process. We proposed the use of cyanocobalamin as an auxiliary substance. It is an electroactive substance due to the presence of cobalt(III) in the corrin ring. The process of vitamin B<sub>12</sub> chemisorption on mesoporous carbon is known [18]. By attaching vitamin B<sub>12</sub> to the surface of the electrode due to adsorption, it is possible to create conditions allowing the binding of methionine from the solution to form an electroactive compound at the electrode. The purpose of this work is to establish the mechanism of methionine oxidation at the modified electrode to optimize the conditions for the determination of methionine by stripping voltammetry.

### *Experimental*

#### *Equipment and reagents*

Methionine (Sigma, USA), multi-walled carbon nanotubes produced by “Graphen” (USA), vitamin B<sub>12</sub> (Belarus) were used in the work. All measurements were taken using standard pH solutions purchased from “Merck” (Germany). The solutions were prepared using deionized water obtained from “Sartorius” of the arium® pro brand. All the experiments were carried out at room temperature. Electrochemical measurements were performed using a TA-LAB voltammetric analyzer (NPO “Tomanalit”) in direct-current mode in a three-electrode cell. The indicator electrode was a modified carbon-containing electrode; silver-chloride electrodes in the 3 M KCl solution were used as an auxiliary electrode and a reference electrode. Multi-walled carbon nanotubes (MCNT) were applied to the surface of the carbon-containing electrode by electrolysis from a MCNT aqueous suspension by means of a universal power source UIP-2.

#### *Modification of the carbon-containing electrode*

MCNT were moistened with a small amount of ethyl alcohol; then water was added. After it MCNT were placed into the ultrasound bath for 3 hours. The ready-to-use suspension was used to apply nanotubes to the surface of the carbon-containing electrode by anodic polarization. A stainless-steel plate was used as the cathode (counter electrode). After 8 s of the electrolysis, the applied MCNT were dried and vitamin B<sub>12</sub> was deposited. Vitamin B<sub>12</sub> was deposited at the electrode modified with MCNT by cycling the potential in the range of  $-1.4 \dots +1.0$  V. The modification of the electrode is described in more detail in [18].

### *Results and Discussion*

The choice of optimal conditions for the oxidation and/ or reduction processes of the substance is the essential task in the development of a quantitative technique. Suitable conditions lead to an increase in the sensitivity of the analysis. The effect of a number of factors, such as the pH of the background electrolyte, accumulation parameters, and the potential sweep rate on the anode signal of methionine, was studied to select the optimal conditions for obtaining an analytical signal.

#### *The effect of pH*

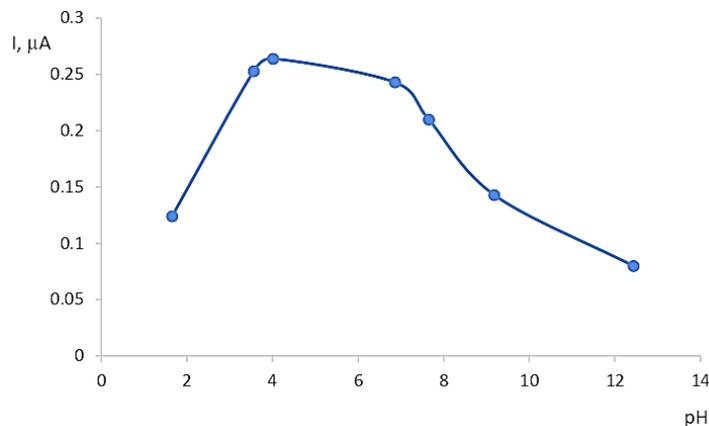
The pH of the background electrolyte has a significant effect on the mechanism of redox processes in voltammetric analysis. Standard buffer solutions from 1.65 to 12.43 were used to establish the optimal value of the hydrogen ion concentration during methionine oxidation at the modified electrode.

Figure 1 shows the dependence of the anode peak value of methionine on the pH of the background electrolyte. With an increase in the pH from 1.65 to 4.01, the peak current of methionine oxidation increases. The decrease in the signal at  $\text{pH} < 4$  is associated with the protonation of the amino groups of methionine and cobalamin; the adsorption of methionine on the electrode is hindered as a result of electrostatic repulsion. The signal intensity decreases when the pH shifts to the neutral and alkaline regions, which is due to the lack

of protons required for methionine oxidation. The peak shift towards more negative values as the pH of the buffer solution increases. A linear dependence can be observed between the peak potential and the pH:

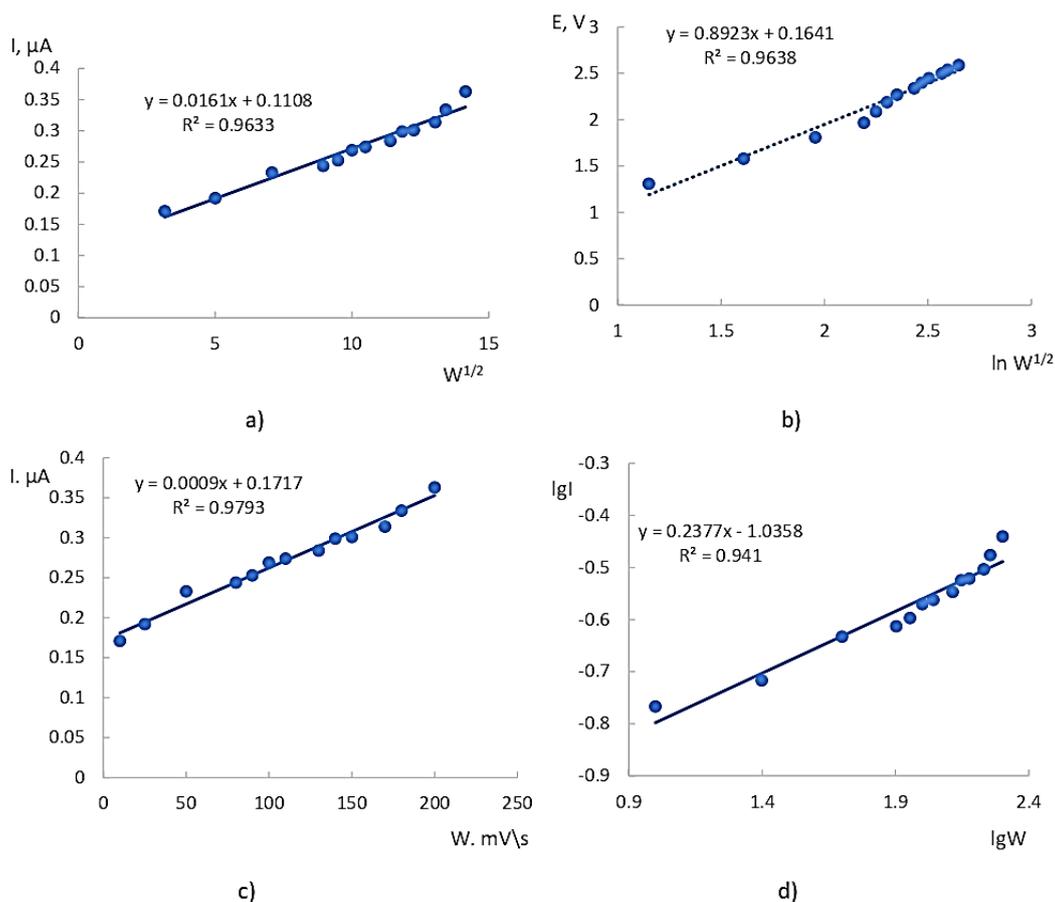
$$Y = -0.0492X + 0.6930. \quad (1)$$

The slope value of 49.2 mV/pH is close to the Nernst slope (59.0 mV/pH), which indicates an equal number of electrons and protons participating in the electrochemical process.



$$E_{cl} = -1.6 \text{ V}; t = 120 \text{ s}; W = 50 \text{ mV/s}; C = 2 \times 10^{-6} \text{ mol/dm}^3$$

Figure 1. Dependence of the electrooxidation current of methionine at the modified electrode on the pH



$$E_{cl} = -1.6 \text{ V}; t = 120 \text{ s}; C = 2 \times 10^{-6} \text{ mol/dm}^3$$

Figure 2. Dependence of the methionine oxidation current on the square root of the potential sweep rate (a), the dependence of the potential on  $\ln W^{1/2}$  (b), the dependences of the current on the potential sweep rate (c) and  $\lg I$  on  $\lg W$  (d)

*The effect of the potential sweep rate on the current and potential of the anode peak*

The dependences of the anode peak current and potential on the potential sweep rate allow us to estimate the mechanism of the electrochemical reaction. In this regard, the influence of the potential sweep rate on the anode current of methionine was studied. To establish the mechanism of methionine electrooxidation at the modified electrode, the dependences of the current on the square root of the sweep rate (Fig. 2a) and the peak potential on  $\ln(W^{1/2})$  were plotted in Figure 2b.

The linear dependence for the current (Fig. 2a) is typical of both reversible and irreversible processes while the linear dependence of the peak potential of methionine on  $\ln W^{1/2}$  evidences the irreversibility of the process (Figure 2b). Figure 2b presents the dependence of the current on the sweep rate and a similar dependence in logarithmic coordinates (Fig. 2d). The linearity of the logarithmic dependence (Fig. 2d) indicates that diffusion is not a limiting stage of the process. The Semerano criterion (the slope of the dependence of  $\lg I$  on  $\lg W$ ) was 0.24, which indicates the absence of the effect of the diffusion process during the oxidation of methionine on the electrode surface. The Semerano criterion should be 0.5 for the processes limited by diffusion [19]. A similar dependence of the current on the sweep rate demonstrates that adsorption is the limiting stage of the process.

The transfer coefficient of the irreversible process (methionine oxidation) was calculated based on the Tafel dependence. The value of the transfer coefficient was 0.44. The obtained value shows that the first electron participates in the limiting stage of the process.

The following formula was used to calculate the electrons in the case of an irreversible process:

$$\left| E_p - E_{p/2} \right| = \frac{47.7}{\alpha n}, \quad (2)$$

where  $E_p$  is the peak potential;  $E_{p/2}$  is the half-peak potential.

The value of  $n = 1.08$  indicates that one electron takes part in the oxidation reaction.

Earlier in [16] we proposed a possible reaction mechanism based on the Raman spectra of pure methionine and its oxidation products from the electrode. Homocysteine has been shown to be the oxidation product of the methionine complex with cyanocobalamin. Based on the study of the above-mentioned kinetics, it is possible to conclude that one electron and one proton participate in the oxidation process of the complex. This may be conditioned by the fact that cobalt, being in the corrin ring of vitamin B<sub>12</sub>, is oxidized to Co(III) during the anodic potential sweep. During cathodic polarization of the electrode, cobalt in cyanocobalamin is first reduced to +2, and then it is the reduced complex that interacts with methionine. The reverse process of cobalt oxidation occurs during anodic sweep: the methyl group of methionine is transferred to cobalamin and passes into the homocysteine solution (Figure 3).

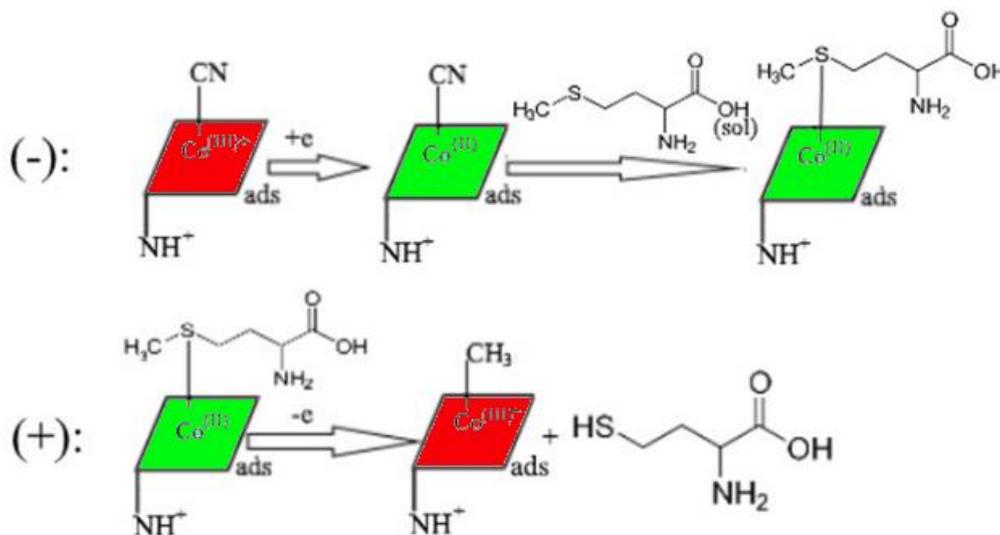


Figure 3. Blank drawing of the methionine oxidation process

*The effect of accumulation parameters on the intensity of the electrooxidation signal*

The range of values from  $-1.9$  V to  $-1.1$  V was studied when choosing the electrolysis potential (Figure 4a). The dependence of the anode peak current on the electrolysis potential passes through the maximum. The anode peaks decrease when the potential is shifted to the negative region, which is most likely due to the release of hydrogen that prevents the formation of the complex on the electrode surface. Therefore, the potential of  $-1.6$  V was chosen as optimal for concentrating methionine at the modified electrode. As can be seen from Figure 4b, after accumulation for more than 300 s, the signal intensity changes slightly, which may be conditioned by the complete filling of the electrode surface with a monolayer of the deposited methionine. Therefore, during subsequent studies, the accumulation was carried out no more than 180 s.

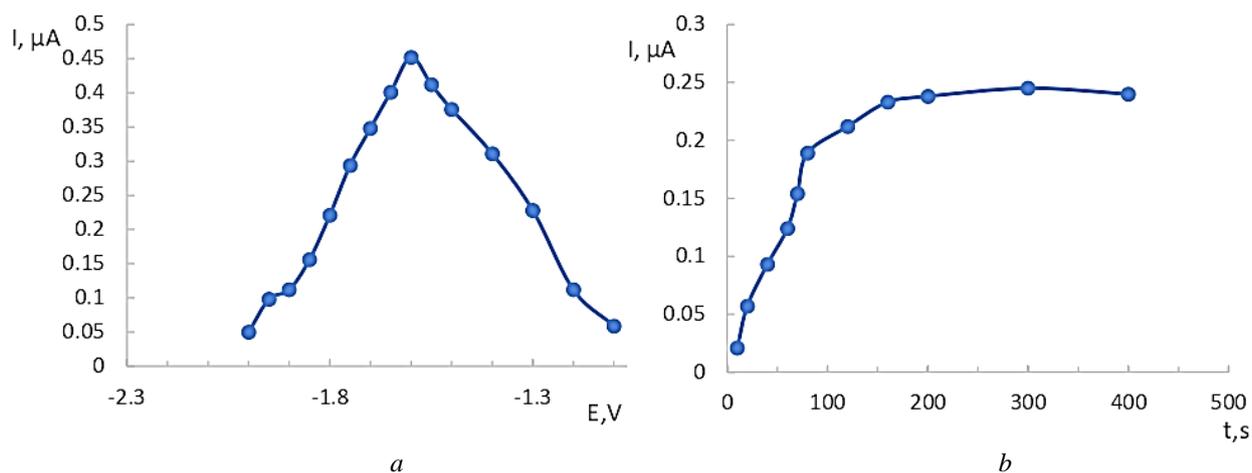


Figure 4. Dependence of the methionine electrooxidation current on the accumulation potential at the modified electrode ( $t = 120$  s,  $W = 80$  mV/s,  $C = 5 \times 10^{-6}$  mol/dm<sup>3</sup>) (a) and the dependence of the methionine electrooxidation current on the accumulation time ( $E_{el} = -1.6$  V,  $W = 80$  mV/s,  $C = 2 \times 10^{-6}$  mol/dm<sup>3</sup>) (b)

Despite the fact that the participation of adsorption during electrooxidation of methionine at the electrode has been established, it can be concluded that the mentioned factor does not prevent quantitative assessment in the specified concentration range. The dependence of current on concentration is linear in the range of  $(1-50) \times 10^{-7}$  and obeys the equation:

$$I = 0.024C + 0.003 \quad (E_{el} = -1.6 \text{ V}, t = 150 \text{ s}, W = 80 \text{ mV/s}). \quad (3)$$

The effect of other amino acids on the results of the determination of methionine was studied. The ultimate transferable concentration was defined as the maximum concentration of a hindrance substance, causing an error in the determination of amino acids of no more than 5 % [19, 20]. The 10-fold excess of ascorbic acid, tryptophan, histidine, glycine, arginine, cysteine and tyrosine does not have a noticeable distorting effect on the procedure for methionine determination.

*Some metrological characteristics of the procedure for tryptophan and 5-hydroxytryptophan determination*

According to ISR 61-2010, the indices of repeatability, intermediate precision, and accuracy were calculated (Table 1).

Table 1

**Metrological characteristics of the procedure for methionine determination ( $p = 0.95$ ,  $n = 2$ ,  $l = 15$ )**

Concentration $\times 10^{-7}$ , mol/dm <sup>3</sup>	Repeatability index, $\sigma_r^*$ , %	Intermediate precision index, $\sigma_{RI}^*$ , %	Accuracy index (relative accuracy limits at $P = 0.95$ ), $\pm \Delta$ , %
1	13.9	15.2	29.0
10	13.3	14.7	28.4
20	12.9	14.3	27.3
30	12.5	13.9	26.9
40	12.0	13.5	25.5
50	11.7	13.3	25.4

Based on the obtained metrological characteristics of the developed procedure for methionine determination, it is possible to conclude that the accuracy index does not exceed 29 %, the repeatability and intermediate precision indices do not exceed 14 % and 16 %, respectively.

The modified electrode makes it possible to obtain stable analytical signals for at least 25 cycles (blank experiment, analyzed assay, spiked sample). Further, the electrode sensitivity decreases and the electrode surface renewal is required. The electrode surface is cleansed by electrolysis for 300 s with a potential of  $-0.5$  V. Then a new film of vitamin B<sub>12</sub> is applied to the surface by cyclic potential scanning.

### Conclusions

One proton and one electron take part in the process of oxidation of methionine complex with cobalamin. During oxidation, methionine takes one proton and one electron. In the process of cathodic polarization, cobalt(III) in cobalamin is reduced to cobalt(II) to which a partially negatively-charged sulfur atom in the methionine molecule is attached due to electrostatic interactions. In case of anodic polarization, cobalt(II) in the complex is oxidized to Co(III), the methyl group is transferred from methionine to cobalamin, while homocysteine passes into solution. The process is controlled by adsorption. The following optimal conditions for methionine determination have been selected: the background electrolyte is the tartrate buffer solution with pH = 4.01; the electrolysis potential is 1.6 V; the accumulation time is up to 180 s. A number of metrological indices have been established for the procedure of methionine determination: accuracy, repeatability and intermediate precision indices. The range of detectable contents was  $(1-50) \times 10^{-7}$ ; the detection limit was  $5.0 \times 10^{-8}$  M.

### References

- 1 Martínez, Y., Li, X., Liu, G., Bin, P., Yan, W., Más, D., Valdivié, M., Hu, C.-A.A., Ren, W., & Yin, Y. (2017). The role of methionine on metabolism, oxidative stress and diseases. *Amino Acids*, 49 (12). 2091–2098. <https://doi.org/10.1007/s00726-017-2494-2>
- 2 Butterfield, D.A. & Boyd-Kimball, D. (2005). The critical role of methionine 35 in Alzheimer's amyloid  $\beta$ -peptide (1-42)-induced oxidative stress and neurotoxicity. *Biochim. Biophys. Acta. Proteins Proteomics*, 1703(2). 149–156. <https://doi.org/10.1016/j.bbapap.2004.10.014>
- 3 Jackson, R. & Hunter, T. (1970). Role of methionine in the initiation of haemoglobin synthesis. *Nature*, 227(5259), 672–676. <https://doi.org/10.1038/227672a0>
- 4 Arber, W. (1965). Host specificity of DNA produced by Escherichia coli: V. The role of methionine in the production of host specificity. *J. Mol. Biol.*, 11(2). 247–256. <https://doi.org/10.1007/BF00268883>
- 5 Reed, D.J. & Orrenius, S. (1977). The role of methionine in glutathione biosynthesis by isolated hepatocytes. *Biochem. Biophys. Res. Commun.*, 77(4). 1257–1264. [https://doi.org/10.1007/978-3-642-67132-6\\_3](https://doi.org/10.1007/978-3-642-67132-6_3)
- 6 Wilcken, D.E. & Wilcken, B. (1976). The pathogenesis of coronary artery disease. A possible role for methionine metabolism. *J. Clin. Invest.*, 57(4). 1079–1082. <http://doi.org/10.1172/JCI108350>
- 7 Man, S. (1960). Polarographie einer Sulfoniumverbindung. *Z. Anal. Chem.*, 173, 112. <https://doi.org/10.1007/BF00448739>
- 8 Lavine, T.F. (1943). An iodometric determination of methionine. *J. Biol. Chem.*, 151. [https://doi.org/10.1016/s0021-9258\(18\)72138-8](https://doi.org/10.1016/s0021-9258(18)72138-8)
- 9 Gensch, K.H. & Higuchi, T. (1967). Kinetic investigation of reversible reaction between methionine and iodine. Improved iodometric determination of methionine. *J. Pharm. Sci.*, 56, 177–184. <https://doi.org/10.1002/jps.2600560205>
- 10 Sysak, P.K., Foote, C.S. & Ching, T.-Y. (1977). Chemistry of singlet oxygen — XXV. Photooxygenation of methionine. *Photochem. Photobiol.*, 26, 19–27. <https://doi.org/10.1111/j.1751-1097.1977.tb07443.x>
- 11 Kyriacou, D. (1966). Rapid voltammetric determination of methionine with a platinum electrode. *Nature*, 211, 519–521. <https://doi.org/10.1038/211519b0>
- 12 Brabec, V. & Mornstein, V. (1980). Electrochemical behaviour of proteins at graphite electrodes. II. Electrooxidation of amino acids. *Biophys. Chem.*, 12, 159–165. [https://doi.org/10.1016/0301-4622\(80\)80048-2](https://doi.org/10.1016/0301-4622(80)80048-2)
- 13 Reynaud, J.A., Malfoy, B. & Canesson, P. (1980). Electrochemical investigations of amino acids at solid electrodes. *J. Electroanal. Chem. Interfacial Electrochem.*, 114, 195–211. [https://doi.org/10.1016/S0022-0728\(80\)80447-5](https://doi.org/10.1016/S0022-0728(80)80447-5)
- 14 Sanaullah, S. Wilson & Glass, R.S. (1994) The effect of pH and complexation of amino acid functionality on the redox chemistry of methionine and X-ray structure of [Co(en)2(L-Met)](ClO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O. *J. Inorg. Biochem.*, 55, 87–99. [https://doi.org/10.1016/0162-0134\(94\)85031-3](https://doi.org/10.1016/0162-0134(94)85031-3)
- 15 Enache, T.A. & Oliveira-Brett, A.M. (2011). Boron doped diamond and glassy carbon electrodes comparative study of the oxidation behaviour of cysteine and methionine. *Bioelectrochemistry*, 81, 46–52. <https://doi.org/10.1016/j.bioelechem.2011.02.001>
- 16 Shelkovnikov, V.V., Altyev, A.M. & Vinogradov, M.E. (2019) Determination of Methionine in Medicines by Stripping Voltammetry. *J. Anal. Chem.*, 74(12), 1239–1244. <https://doi.org/10.1134/S1061934819120116>

17 Manta, B. & Relover, S. (2017). Regulated methionine oxidation by monooxygenases. *Free Radical Biology and Medicine*, 109, 141. <https://doi.org/10.1016/j.freeradbiomed.2017.02.010>

18 Jose, A. & Lima, Jr. (2013). Using Raman spectroscopy to understand the origin of the phase transition observed in the crystalline sulfur based amino acid l-methionine. *Vibrational Spectroscopy*, 65, 132–135. <https://doi.org/10.1016/j.vibspec.2012.12.004>

19 Keyvanfar, M., Shakeri, R., Karimi-Maleh, H. & Alizad, K. (2013). Highly selective and sensitive voltammetric sensor based on modified multiwall carbon nanotube paste electrode for simultaneous determination of ascorbic acid, acetaminophen and tryptophan. *Mater. Sci. Eng. C*, 33, 811-816. <https://doi.org/10.1016/j.msec.2012.11.005>

20 Rajabzadeh, N., Benvidi, A., Mazloum-Ardakani, M., Firouzabadi, A.D. & Vafazadeh, R. (2015). A Highly Sensitive Sensor Based on Reduced Graphene Oxide. Carbon Nanotube and a Co (II) Complex Modified Carbon Paste Electrode: Simultaneous Determination of Isoprenaline. Captopril. Tryptophan. *Electroanal.*, 27, 2792–2798. <https://doi.org/10.1002/elan.201500236>

В.В. Шелковников, А.М. Алтыев, М.С. Фрянова

## В<sub>12</sub> дәрумені мен көпқабырғалы көміртекті нанотүтікшелермен модификацияланған электродта метиониннің электр тотығын зерттеу

Жұмыста В<sub>12</sub> витаминімен және көпқабырғалы көміртекті нанотүтіктермен модификацияланған электродтағы метиониннің электрохимиялық тәртібі зерттелген. Модификацияланған электродта болатын тотығу-тотықсыздану процестерінің ықтимал механизмі ұсынылған. Электродты қалыптастыру кезінде В<sub>12</sub> дәрумені адсорбция есебінен көпқабатты көміртекті нанотүтікшелермен модификацияланған көміртегі бар электродтың бетіне бекітіледі. Катодты поляризация кезінде кобаламиндегі кобальт (III) кобальт (II) дейін азаяды, оған электростатикалық өзара әрекеттесулер есебінен метионин молекуласындағы жартылай теріс зарядталған күкірт атомы қосылады. Анодты поляризация кезінде кешендегі кобальт (II) Со<sup>3+</sup> дейін тотығады, метиониннен метил тобы кобаламинге ауысады, бұл ретте гомоцистеин ерітіндіге өтеді, яғни бұл комбинациялық шашырау спектрлерімен дәлелденген. Ток пен потенциалдың жаймалау жылдамдығына тәуелділігін зерттеу негізінде тотығу процесі қайтымсыз екендігі анықталды. Шектеу сатысы адсорбция болып табылады, ал бір протон мен бір электрон электрохимиялық сатыға қатысады. Аналитикалық сигналды тіркеудің келесі оңтайлы шарттары таңдалды: фондық электролит-тарtratтық буферлік ерітінді рН = 4,01; электролиз потенциалы — 1,6 В; жинақтау уақыты — 180 с дейін. Метионинді анықтау әдісінің метрологиялық сипаттамаларына бағалау жүргізілді. Дәлдік көрсеткіші 29 %-дан аспайды. Қайталану және аралық дәлдік көрсеткіштері сәйкесінше 14% және 16% құрады. Анықталған құрамдардың диапазоны (1–50)×10<sup>-7</sup>; анықтау шегі — 5,0×10<sup>-8</sup> М. Аскорбин қышқылының, триптофанның, глициннің, цистеиннің және тирозиннің 10 есе артық болуы метионинді анықтау әдісіне айтарлықтай бұрмалаушы әсер етпейтіні көрсетілген.

*Клт сөздер:* метионин, модификацияланған электрод, В<sub>12</sub> витамині, электр тотығу, кобаламин, электрон, протон, инверсиялы вольтамперметрлеу, көпқабырғалы көміртекті нанотүтіктер.

В.В. Шелковников, А.М. Алтыев, М.С. Фрянова

## Изучение процесса электроокисления метионина на модифицированном витамином В<sub>12</sub> и многостенными углеродными нанотрубками электроде

В статье изучено электрохимическое поведение метионина на электроде, модифицированном витамином В<sub>12</sub> и многостенными углеродными нанотрубками. Предложен возможный механизм окислительно-восстановительных процессов, протекающих на модифицированном электроде. При формировании электрода витамин В<sub>12</sub> закрепляется на поверхности углеродсодержащего электрода модифицированного многостенными углеродными нанотрубками за счет адсорбции. При катодной поляризации кобальт (III) в кобаламине восстанавливается до кобальта (II), на который, за счет электростатических взаимодействий, присоединяется частично отрицательно заряженный атом серы в молекуле метионина. При анодной поляризации кобальт (II) в комплексе окисляется до Со<sup>3+</sup>, метильная группа с метионина переносится на кобаламин, при этом в раствор переходит гомоцистеин, что доказано спектрами комбинационного рассеяния. На основании изучения зависимости тока и потенциала от скорости развертки было установлено, что процесс окисления необратим. Лимитирующей стадией является адсорбция, а в электрохимической стадии участвуют один протон и один электрон. Были подобраны следующие оптимальные условия регистрации аналитического сигнала: фоновый электролит — тарtratный буферный раствор рН = 4,01; потенциал электролиза — 1,6 В; время накопления — до 180 с. Проведена оценка метрологических характеристик методики определения метионина. Показатель точности не превышал 29 %. Показатели повторяемости и промежуточной прецизионности не превы-

шали 14 и 16 % соответственно. Диапазон определяемых содержаний составил  $(1-50) \times 10^{-7}$ ; предел обнаружения —  $5,0 \times 10^{-8}$  М. Показано, что 10-кратный избыток аскорбиновой кислоты, триптофана, глицина, цистеина и тирозина не оказывает заметного искажающего влияния на методику определения метионина.

*Ключевые слова:* метионин, модифицированный электрод, витамин В<sub>12</sub>, электроокисление, кобаламин, электрон, протон, инверсионная вольтамперометрия, многостенные углеродные нанотрубки.

#### Information about authors\*

**Shelkovnikov, Vladimir Vitalievich** (*corresponding author*) — Candidate of Chemical Sciences Do-  
cent, Head of the Department of Analytical Chemistry, National Research Tomsk State University, Lenin  
avenue, 36, 634050, Tomsk, Russia, e-mail: [shvv@chem.tsu.ru](mailto:shvv@chem.tsu.ru); <https://orcid.org/0000-0002-5778-2862>

**Altiev, Alexey Muratovich** — postgraduate student of the Faculty of Chemistry, National Research  
Tomsk State University, Lenin avenue, 36, 634050 Tomsk, Russia, e-mail: [lexa-cors@mail.ru](mailto:lexa-cors@mail.ru);  
<https://orcid.org/0000-0001-8543-3451>

**Fryanova, Maria Sergeevna** — student of the Faculty of Chemistry, National Research Tomsk State  
University, Lenin avenue, 36, 634050 Tomsk, Russia, e-mail: [m.s.fryanova@mail.ru](mailto:m.s.fryanova@mail.ru); <https://orcid.org/0000-0002-6394-2498>

---

\*The author's name is presented in the order: *Last Name, First and Middle Names*